



Concise access to enantiopure (*S*)- and (*R*)- α -trifluoromethyl pyroglutamic acids from ethyl trifluoropyruvate-based chiral CF₃-oxazolidines (Fox)

Grégory Chaume^a, Marie-Céline Van Severen^a, Louis Ricard^b, Thierry Brigaud^{a,*}

^a Laboratoire de Synthèse Organique Sélective et Chimie Organométallique (SOSCO), UMR CNRS 8123, Université de Cergy-Pontoise, 5, Mail Gay Lussac, Neuville sur Oise, 95031 Cergy-Pontoise Cedex, France

^b Laboratoire Hétéroéléments et Coordination, UMR CNRS 7653, Ecole Polytechnique, 91128 Palaiseau, France

ARTICLE INFO

Article history:

Received 17 June 2008

Received in revised form 18 July 2008

Accepted 24 July 2008

Available online 3 August 2008

Keywords:

α -Trifluoromethyl amino acids

Oxazolidines

Organofluorine chemistry

Stereoselective synthesis

Ring closure

Lactams

ABSTRACT

A straightforward synthesis of enantiopure (*S*)- and (*R*)- α -Tfm-pyroglutamic acid is reported. The strategy is based on the use of a chiral CF₃-hydroxymorpholinone intermediate conveniently obtained from ethyl trifluoropyruvate-based chiral CF₃-oxazolidines (Fox). The key step is an oxidative cyclization followed by a reductive cleavage of the (*R*)-phenylglycinol chiral auxiliary.

© 2008 Elsevier B.V. All rights reserved.

1. Introduction

Conformationally constrained cyclic amino acids have recently gained considerable interest because of their ability to control the conformation of peptides for structure–activity relationships investigations as well as for the design of peptidomimetics [1]. In particular, incorporation of a proline unit is known to restrict the amino acyl-proline *cis/trans* isomerization [2], to limit the protein folding and consequently to modulate the biological activity of peptides. Among the numerous proline derivatives reported in the literature, fluorinated proline-type amino acids have received increasing attention [3]. This is particularly due to the unique physical and biological properties induced by the introduction of fluorinated groups in peptides and peptidomimetics [4]. However, their use in peptide chemistry remains very limited due to the difficulty to prepare them efficiently, particularly in their enantiopure form. Pyroglutamic acid derivatives were also reported to be efficient tools for the control of the peptidyl bond geometry [5]. Although several examples of fluorine containing pyroglutamic derivatives are reported in the literature [6], the

preparation of α -Tfm-pyroglutamic acids in enantiopure form has never been reported [7].

In the course of our studies, we recently reported several approaches for the stereoselective synthesis of α - and β -trifluoromethyl amino acids (Tfm AAs) starting from chiral CF₃-oxazolidines (Fox) or imines [8]. Among them, we developed an efficient route for the synthesis of (*S*)- and (*R*)- α -Tfm proline based on the use of the key chiral CF₃-hydroxymorpholinone **2** (Scheme 1) [9]. This compound was conveniently prepared in a few step from the oxazolidines **1** derived from ethyl trifluoropyruvate.

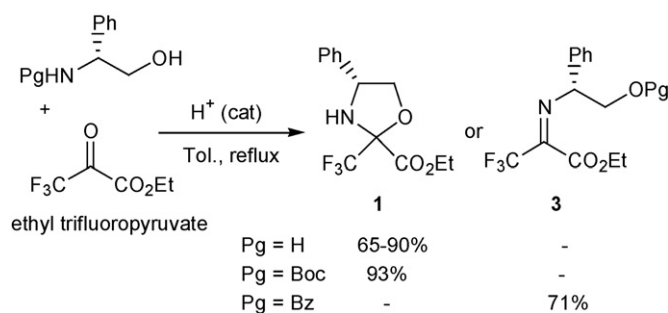
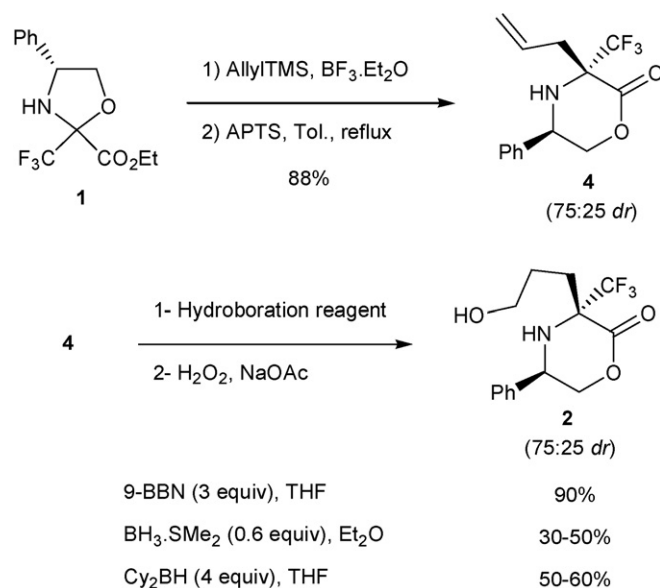
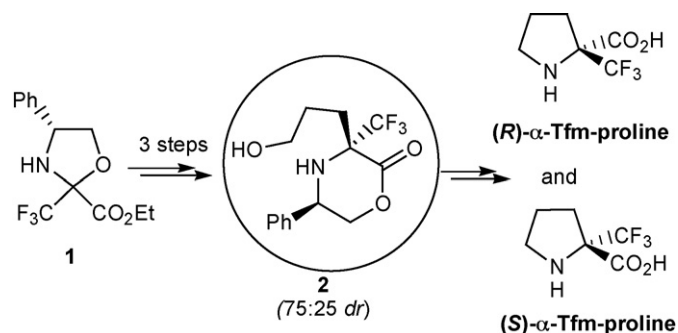
We present here another feature of the use of the CF₃-hydromorpholinone **2**. This versatile intermediate proved to be also highly valuable for the synthesis of α -Tfm-pyroglutamic acids in enantiopure form.

2. Results and discussion

The preparation of the starting CF₃-hydroxymorpholinone **2** was based on our reported procedure [9]. In addition, the first step involving the formation of oxazolidines **1** from ethyl trifluoropyruvate and (*R*)-phenylglycinol was significantly improved. We observed that the yield of the direct condensation was not entirely reproducible (65–90%) [10]. This was probably due to both bis-nucleophilic reactivity of (*R*)-phenylglycinol and bis-electrophilic reactivity of ethyl trifluoropyruvate. In order to increase the

* Corresponding author. Fax: +33 1 34 25 73 81.

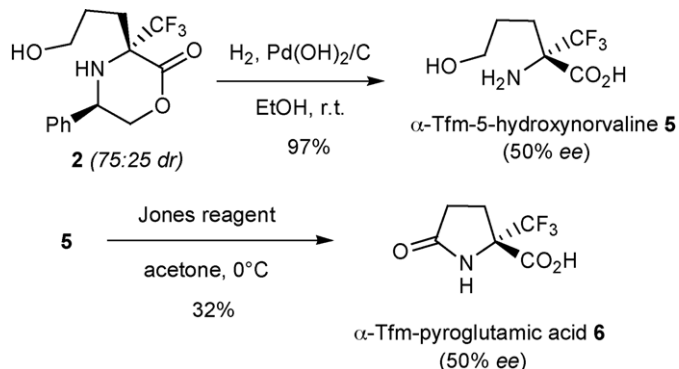
E-mail address: thierry.brigaud@u-cergy.fr (T. Brigaud).

**Scheme 2.**

selectivity of the oxazolidinone **1** formation, we designed to use a *N*-protected (*R*)-phenylglycinol which would give an intermediate ethyl trifluoropyruvate hemiacetal. After removal of the protecting group, the cyclization of the hemiacetals would lead to the expected oxazolidinones. We were pleased to observe that the reaction of the *N*-Boc (*R*)-phenylglycinol under acidic catalysis (0.1 equiv. PPTS) gave directly the oxazolidinones **1** (75:25 diastereomeric mixture) in a completely reproducible high yield (93%) (Scheme 2). Intriguingly, when reaction was performed starting from the *N*-Bz phenylglycinol, the only corresponding *O*-Bz imine **3** was isolated in 71% yield. We postulate that in both cases, there is a protecting group transfer from the amino to the hydroxyl group of the (*R*)-phenylglycinol in the acidic reaction conditions. The free amino group would then react selectively with the carbonyl group of the ethyl trifluoropyruvate to give an imine intermediate. The reaction stopped at the imine stage **3** when the stable benzoyl protecting group was used. However, with the Boc protecting group, the acidic mediated smooth removal of the Boc group should undergo a selective intramolecular cyclization of the resulting hydroxyimine into oxazolidinones **1**.

The CF₃-hydroxymorpholinone **2** was then obtained as a non-separable 75:25 diastereomeric mixture following our previously reported three-step allylation/lactonisation/hydroboration sequence from oxazolidinones **1** (Scheme 3) [9]. The hydroboration reaction of **4** was performed in high yield (90%) using 9-BBN. Lowest yield were achieved using BH₃·SMe₂ or dicyclohexylborane (30–60%).

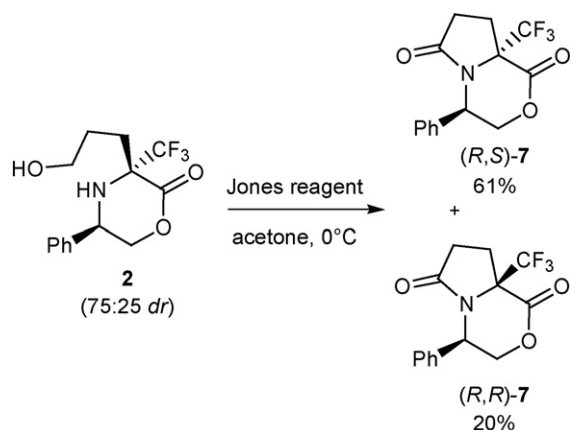
As an exploratory study, the synthesis of the α -Tfm-pyroglutamic acid **6** from the 75:25 diastereomeric mixture of the hydroxymorpholinone **2** was investigated. The target compound was obtained by the following sequence involving the removal of the chiral auxiliary followed by the oxidation of the hydroxyl group into the corresponding acid and the ring closure by lactamization (Scheme 4). The hydrogenolysis of the hydroxymorpholinone **2** catalyzed by Pearlman's catalyst led to the new α -Tfm-5-hydroxynorvaline **5** in 97% yield. Finally, the expected α -Tfm-



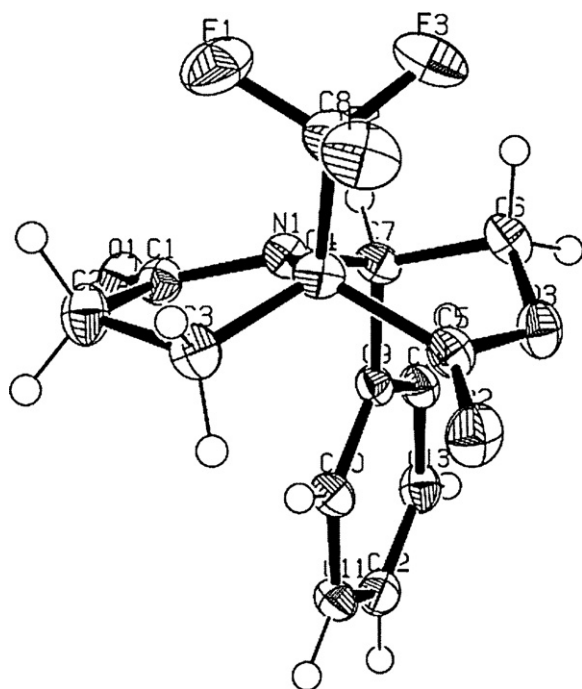
pyroglutamic acid **6** (50% *ee*) was obtained by oxidative cyclization using Jones reagent in 32% non-optimized yield. The direct cyclization of the intermediate amino acid is in accordance with the observation of Burger et al. [7] since they reported in the racemic series that α -Tfm-glutamic acid gave rise to spontaneous cyclization into α -Tfm-pyroglutamic acid.

The main drawback of this synthetic pathway is that it supposes an efficient separation of both diastereomers of the CF₃-hydroxymorpholinone **2** in order to obtain enantiopure α -Tfm-pyroglutamic acids. As the chromatographic separation of both diastereomers of **2** is difficult, we anticipated that the separation of the bicyclic lactams resulting from oxidative cyclization of **2** would be more convenient [11]. Hence the diastereomeric mixture of **2** was subjected to oxidation using the convenient Jones reagent to give the corresponding bicyclic lactams **7** in high yield (Scheme 5) [12].

As expected, at this stage, both diastereomers of the bicyclic lactams **7** were very easily separated by silica gel chromatography to afford (*R,S*)-**7** in 61% isolated yield and (*R,R*)-**7** in 20% isolated yield (Scheme 5) [13]. Moreover, we were pleased to observe that both diastereomers could also be very conveniently separated by selective crystallization and filtration. Indeed, (*R,S*)-**7** is a white solid poorly soluble in diethyl ether, whereas (*R,R*)-**7** is an oil completely soluble in this solvent. This very efficient and convenient separation of both diastereomers will provide an easy

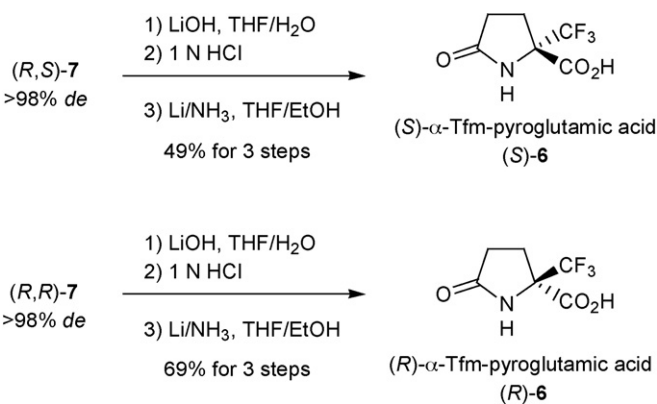


Scheme 5.

Fig. 1. ORTEP view of **(R,S)-7**.

access to enantiopure derivatives of **7** in large scale. The **(R,S)** configuration assignment of the major diastereomer of **7**, was confirmed by single crystal X-ray analysis (Fig. 1) [14].

The removal of the **(R)**-phenylglycinol chiral auxiliary was then considered. In accordance with the observations reported by Ma et al. [15], all attempts to cleave the *N*-benzylic bond of the diastereomerically pure **(R,S)-7** by Pd/C or Pd(OH)₂/C catalyzed hydrogenolysis under acidic or neutral conditions failed. However, the removal of the chiral auxiliary was suitably achieved in a three-step procedure involving the saponification of the lactone ring by LiOH, acidification with 1N HCl followed by a reductive cleavage of the benzylic bond using lithium in liquid ammonia [15,16]. Following this procedure, enantiopure **(S)**- α -Tfm-pyrroglutamic acid **(S)-6** was obtained in 49% yield from **(R,S)-7**. The enantiopurity (>98% ee) of **(S)-6** was confirmed by its derivatization into the corresponding diastereomerically pure **(S)**-1-phenylethylamide. In a similar manner, **(R)**- α -Tfm-pyrroglutamic acid **(R)-6** was obtained in enantiopure form in 69% yield from **(R,S)-7** (Scheme 6).



Scheme 6.

3. Conclusion

In conclusion, we have successfully developed a concise synthesis of both enantiomers of the highly constrained α -Tfm-pyrroglutamic acid in enantiopure form from a highly versatile chiral CF₃-hydroxymorpholinone intermediate. Further investigations about synthetic applications of this conformationally constrained cyclic amino acid are underway and will be reported in due course.

4. Experimental

General: Unless otherwise mentioned, all the reagents were purchased from commercial source. THF was distilled under nitrogen from sodium/benzophenone prior to use. ¹H NMR (400.00 MHz), ¹³C NMR (100.50 MHz) and ¹⁹F NMR (376.20 MHz) were measured on a JEOL 400 spectrometer. Chemical shifts of ¹H NMR were expressed in parts per million downfield from tetramethylsilane ($\delta = 0$) in CDCl₃. Chemical shifts of ¹³C NMR were expressed in parts per million downfield from CDCl₃ as internal standard ($\delta = 77.0$). Chemical shifts of ¹⁹F NMR were expressed in parts per million downfield from C₆F₆ as internal standard ($\delta = -164.9$). Coupling constants are reported in hertz. Column chromatography was performed on Merck Kieselgel 60 (0.040–0.063 mm), employing mixture of specified solvent as eluent. Thin-layer chromatography (TLC) was performed on Merck silica gel (Merck 60 PF₂₅₄) plates. Silica TLC plates were visualized under UV light, by a 10% solution of phosphomolybdic acid in ethanol followed by heating. Mass spectra (MS) were obtained on a GC/MS apparatus HP 5973 MSD with an HP 6890 Series GC. Ionization was obtained by electronic impact (EI 70 eV). Infrared spectra (IR) were obtained by Fourier-transformation on BRÜCKER TENSOR 27, wavenumbers are given in cm⁻¹. Elemental analyses were performed by the CNRS analysis central service. Optical rotations are reported as their specific rotations determined using a JASCO DIP-370 polarimeter. Melting points were obtained on a Büchi apparatus and are uncorrected.

4.1. (4R)-2-Ethoxycarbonyl-2-trifluoromethyl-4-phenyl-1,3-oxazolidine (**1**) [9]

To a stirred solution of 5.14 g of **(R)**-*N*-Boc-phenylglycinol (21.7 mmol, 1.0 equiv.) in 200 mL of toluene at room temperature were added 1.09 g of pyridinium *p*-toluenesulfonate (4.34 mmol, 0.2 equiv.) and 3.17 mL of ethyl trifluoropyruvate (23.8 mmol, 1.1 equiv.). The mixture was stirred for 1 h at room temperature, warmed to 140 °C with a Dean–Stark apparatus for 20 h, and then

cooled to 0 °C with an ice-bath. The resulting mixture was filtered and toluene was evaporated. Purification by flash chromatography (95:5 cyclohexane/ethyl acetate) gave 5.80 g (93%) of **1** (75:25 diastereomeric mixture).

4.2. (*E*)-1-Ethoxycarbonyl-2,2,2-trifluoroethylidene (1*R*)-2-benzoyloxy-1-phenylethylamine (**3**)

To a stirred solution of 200 mg of (*R*)-*N*-Bz-phenylglycinol (0.83 mmol, 1.0 equiv.) in 10 mL of toluene at room temperature, were added 23 mg of *p*-toluenesulfonic acid (0.12 mmol, 0.15 equiv.) and 110 μ L of ethyl trifluoropyruvate (0.83 mmol, 1.0 equiv.). The mixture was warmed to 140 °C with a Dean–Stark apparatus for 20 h, and then cooled to 0 °C with an ice-bath. The resulting mixture was filtered and toluene was evaporated. Purification by flash chromatography (9:1 cyclohexane/ethyl acetate) gave 229 mg of imine **3** (71%). Colorless oil; $[\alpha]_D^{20}$ -3.05 (c 2.1, CHCl₃); IR (neat): 2987, 1742, 1684, 1452, 1272, 713 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ = 1.19 (t, *J* = 7.1 Hz, 3H), 4.17 (dq, *J* = 10.8, 7.1 Hz, 1H), 4.30 (dq, *J* = 10.8, 7.1 Hz, 1H), 4.50 (dd, *J* = 11.1, 9.3 Hz, 1H), 4.74 (dd, *J* = 11.1, 3.8 Hz, 1H), 5.33 (dd, *J* = 9.3, 3.8 Hz, 1H), 7.37–7.47 (m, 8H), 8.02–7.99 (m, 2H); ¹³C NMR (100.5 MHz, CDCl₃): δ = 13.8, 62.9, 66.3, 67.8, 118.1 (q, *J* = 279.0 Hz), 127.3, 128.4, 128.6, 128.9, 129.6, 133.2, 136.8, 150.2 (q, *J* = 36.4 Hz), 158.4, 166.0; ¹⁹F NMR (376.2 MHz, CDCl₃): δ = -72.9 (s); MS (EI) *m/z* 393 [M⁺], 271, 243, 198, 105 (100), 77; Anal. Calcd for C₂₀H₁₈F₃NO₄: C, 61.07; H, 4.61; N, 3.56. Found: C, 61.01; H, 5.01; N, 3.45.

4.3. α -Trifluoromethyl-5-hydroxynorvaline (**5**)

To a 75:25 diastereomeric mixture of hydroxymorpholinone **2** (207 mg, 0.68 mmol) in EtOH (20 mL) was added 380 mg of 20% Pd(OH)₂/C. The reaction was stirred for 48 h under 1 bar atmosphere pressure of hydrogen. The resulting mixture was filtered and evaporated under reduced pressure. The crude mixture was taken up with ether and water. The aqueous phase was washed with ether and evaporated to afford 133 mg of **5** (97%) as white solid. ¹H NMR (400 MHz, D₂O): δ = 1.21–1.33 (m, 1H), 1.34–1.49 (m, 1H), 1.80 (ddd, *J* = 16.7, 11.9, 5.0 Hz, 1H), 2.03 (ddd, *J* = 16.7, 11.9, 4.6 Hz, 1H), 3.39 (td, *J* = 6.0, 2.8 Hz, 2H); ¹³C NMR (100.5 MHz, D₂O): δ = 27.7, 29.4, 63.1, 68.2 (q, *J* = 26.8 Hz), 125.9 (q, *J* = 283.7 Hz), 169.3; ¹⁹F NMR (376.2 MHz, D₂O): δ = -76.2 .

4.4. 8 α -Trifluoromethyl-4-phenyltetrahydropyrrolo[2,1-*c*][1,4]oxazine-1,6-dione (**7**)

To a stirred solution of 263 mg of a 75:25 diastereomeric mixture of alcohol **2** (0.87 mmol, 1.0 equiv.) in 2.8 mL of acetone was added dropwise at 0 °C 634 μ L of solution of Jones reagent (2.74 M in 3.2:1 H₂O/H₂SO₄ solution, 1.7 mmol, 2.0 equiv.). After 20 min at 0 °C, 250 μ L of solution of Jones reagent was added once more and the reaction mixture was stirred for another 20 min at 0 °C. Finally, isopropanol was added until color turned green and the resulting solution was diluted in 50 mL of diethyl ether and 25 mL of water. The layers were separated and the aqueous phase was extracted with AcOEt (3 mL \times 20 mL). The combined organic extracts were washed with saturated aqueous NaHCO₃ solution, dried over MgSO₄ and evaporated under reduced pressure. Purification by flash chromatography (9:1 cyclohexane/ethyl acetate) gave 157 mg (61%) of the diastereomer (*R,S*)-**7** as a white solid and 53 mg (20%) of the diastereomer (*R,R*)-**7** as a colorless oil. (*R,S*)-**7**: white solid; m.p.: 189–190 °C; *R_f* = 0.41 (7:3 cyclohexane/ethyl acetate);

$[\alpha]_D^{25}$ -124.4 (c 0.26, CHCl₃); IR (neat): 2360, 1763, 1725, 1226, 1172, 1151, 701 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ = 2.34–2.48 (m, 1H), 2.60–2.74 (m, 2H), 2.86–2.98 (m, 1H), 4.40 (dd, *J* = 11.5, 1.4 Hz, 1H), 5.05 (dd, *J* = 11.5, 4.1 Hz, 1H), 5.07 (dd, *J* = 4.1, 1.4 Hz, 1H), 7.04–7.08 (m, 2H), 7.27–7.40 (m, 3H); ¹³C NMR (100.5 MHz, CDCl₃): δ = 28.6, 29.2, 53.0, 66.6 (q, *J* = 29.7 Hz), 71.4, 124.3 (q, *J* = 288.9 Hz), 125.6, 128.5, 129.2, 138.2, 165.3, 174.8; ¹⁹F NMR (376.2 MHz, CDCl₃): δ = -76.67 (s); MS (EI) *m/z* 299 [M⁺], 230, 212, 104 (100), 91; Anal. Calcd for C₁₄H₁₂F₃NO₃: C, 56.12; H, 4.04; N, 4.68. Found: C, 56.03; H, 4.06; N, 4.61. (*R,R*)-**7**: colorless oil; *R_f* = 0.29 (7:3 cyclohexane/ethyl acetate); $[\alpha]_D^{25}$ -1.6 (c 0.8, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ = 2.40 (ddd, *J* = 16.0, 9.2, 1.4 Hz, 1H), 2.60–2.78 (m, 2H), 2.87–2.92 (m, 1H), 4.49 (t, *J* = 12.8 Hz, 1H), 4.59 (ddd, *J* = 12.8, 6.9, 1.4 Hz, 1H), 5.04 (dd, *J* = 12.8, 6.9 Hz, 1H), 7.26–7.30 (m, 2H), 7.32–7.45 (m, 3H); ¹³C NMR (100.5 MHz, CDCl₃): δ = 28.7, 28.8, 53.7, 66.3 (q, *J* = 30.1 Hz), 68.8, 124.2 (q, *J* = 284.9 Hz), 127.2, 128.5, 129.1, 134.8, 165.7, 176.6; ¹⁹F NMR (376.2 MHz, CDCl₃): δ = -76.75 (s); MS (EI) *m/z* 299 [M⁺], 230, 212, 104 (100).

4.5. (*S*)- α -Trifluoromethylpyroglutamic acid ((*S*)-**6**)

To a solution of pure bicyclic diastereomer (*R,S*)-**7** (372 mg, 1.04 mmol) in THF (6.3 mL) was slowly added at 0 °C, a 1 M aqueous solution of LiOH (1.25 mL, 1.25 mmol, 1.2 equiv.). The resulting mixture was vigorously stirred at 0 °C for 2 h, and then diluted with ether (10 mL). The layers were separated and the organic layer was washed with water (2 \times 10 mL). The combined aqueous layers were then acidified with 1N HCl until pH 2, extracted with EtOAc (3 \times 10 mL), dried over Na₂SO₄ and concentrated to give 380 mg of crude acid directly used in the next step without further purification.

A solution of the above acid (1.04 mmol) in anhydrous THF (25 mL) and absolute ethanol (2.5 mL) was added to an oven dried three-neck round bottom flask charged with argon. The flask was cooled to -40 °C bath, liquid NH₃ (approximately 150 mL) was condensed using an acetone/liquid nitrogen cold trap flask. Fresh polished lithium flakes were added to the reactants at -40 °C until the deep blue color was kept for 5 min. The reaction was quenched by the addition of NH₄Cl powder (2.7 g), and the reaction was left in the hood until almost all the NH₃ evaporated. The reaction mixture was then diluted with EtOAc (15 mL) and water (15 mL). The aqueous layer was washed with EtOAc (2 \times 20 mL), acidified with 1N HCl until pH 2 and extracted with EtOAc (3 \times 20 mL). The combined organic layers were evaporated to afford 100 mg of pure (*S*)-**6** (49%). White solid; m.p.: 143–144 °C; $[\alpha]_D^{22}$ -12.6 (c 2.2, MeOH); IR (neat): 3139, 3049, 1670, 1402, 1155 cm⁻¹; ¹H NMR (400 MHz, CD₃OD): δ = 2.35–2.55 (m, 4H); ¹H NMR (400 MHz, D₂O): δ = 2.20–2.40 (m, 4H); ¹³C NMR (100.5 MHz, CD₃OD): δ = 26.9; 30.3; 69.0; 126.2 (q, *J* = 283.6 Hz), 170.2, 180.3; ¹³C NMR (100.5 MHz, D₂O): δ = 25.3, 29.0, 67.8 (q, *J* = 28.8 Hz), 124.2 (q, *J* = 283.6 Hz), 170.4, 181.3; ¹⁹F NMR (376.2 MHz, D₂O): δ = -79.0 (s); MS (EI) *m/z* 152 (M⁺–CO₂H) (100); Anal. Calcd for C₆H₆F₃NO₃: C, 36.56; H, 3.07; N, 7.11. Found: C, 36.93; H, 3.10; N, 6.97.

4.6. (*R*)- α -Trifluoromethylpyroglutamic acid ((*R*)-**6**)

According to the procedure described for (*S*)-**6**, starting from (*R,R*)-**7** (551 mg, 1.84 mmol) in THF (11 mL) and 1 M aqueous solution of LiOH (2.21 mL, 2.21 mmol, 1.2 equiv.), crude acid (451 mg) was obtained. This acid was directly used in the next reductive debenzoylation step without further purification to afford 222 mg of pure (*R*)-**6** (69%). White solid; $[\alpha]_D^{25}$ $+11.7$ (c 2.2, MeOH). The spectral data of (*R*)-**6** were identical to those of (*S*)-**6**.

4.7. (*S,S*)-5-oxo-2-trifluoromethylpyrrolidine-2-carboxylic acid (1-phenylethyl)amide (*S,S*)-**8**)

To a solution of (*S*)-phenylethylamine (69 μL , 0.53 mmol, 1.5 equiv.) in DMF (1.5 mL) was successively added at 0 °C Et_3N (203 μL , 1.45 mmol, 4.1 equiv.), HOBt (72 mg, 0.53 mmol, 1.5 equiv.), EDCI (102 mg, 0.53 mmol, 1.5 equiv.) and pure (*S*)-**6** (70 mg, 0.36 mmol) in DMF (1 mL). The resulting mixture was stirred at 0 °C for 20 min and then at room temperature overnight. Then, the reaction mixture was diluted with AcOEt (20 mL) and washed with 1N HCl (10 mL). The layers were separated and the aqueous layer was extracted with EtOAc (2×10 mL). The combined organic layers were then washed with saturated aqueous NaHCO_3 solution (10 mL) and water (10 mL), dried over MgSO_4 and evaporated under reduced pressure. The ^1H and ^{19}F NMR analysis of the crude mixture confirmed that one single diastereomer was obtained. Purification by flash chromatography (1:1 cyclohexane/ethyl acetate) gave 43 mg (51%) of the diastereomer (*S,S*)-**8** as a colorless oil. ^1H NMR (400 MHz, CDCl_3): δ = 1.49 (d, J = 6.9 Hz, 3H), 2.17 (ddd, J = 14.5, 9.2, 4.8 Hz, 1H), 2.30–2.40 (m, 1H), 2.40–2.55 (m, 2H), 5.10 (dq, J = 7.8, 6.9 Hz, 1H), 7.20–7.35 (m, 5H), 7.47 (d, J = 7.8 Hz, 1H), 7.87 (s, 1H); ^{13}C NMR (100.5 MHz, CDCl_3): δ = 21.4, 26.0, 29.0, 49.7, 67.5 (q, J = 28.8 Hz); 124.6 (q, J = 285.6 Hz), 126.1, 127.6, 128.6, 142.3, 165.3, 178.6; ^{19}F NMR (376.2 MHz, CDCl_3): δ = –80.1 (s).

Acknowledgements

The authors thank Ingrid Nativel for her experimental contribution on the hydroboration reaction using dicyclohexylborane. The authors are also grateful to Central Glass Company for their financial support and their generous gift of ethyl trifluoropyruvate.

Appendix A. X-ray crystallographic study

Data were collected at 150.0(1)K on a Nonius Kappa CCD diffractometer using a Mo $K\alpha$ (λ = 0.71070 Å) X-ray source and a graphite monochromator. All data were measured using phi and omega scans. The crystal structures were solved using SIR9744 and Shelxl-97. CCDC-695212 contains the supplementary crystallographic data for this paper. These data can be obtained free of charge at www.ccdc.cam.ac.uk/conts/retrieving.html or from the Cambridge Crystallographic Data Centre, 12, Union Road, Cambridge CB2 1EZ, UK; fax: (internat.) + 44 1223/336 033; E-mail: deposit@ccdc.cam.ac.uk

Compound: (*R,S*)-8a-Trifluoromethyl-4-phenyltetrahydropyrrolo[2,1-c][1,4]oxazine-1,6-dione.

Molecular formula	$\text{C}_{14}\text{H}_{12}\text{F}_3\text{N}_3\text{O}_3$
Molecular weight	299.25
Crystal habit	Block colorless
Crystal dimensions (mm)	$0.20 \times 0.18 \times 0.18$
Crystal system	Monoclinic
Space group	$P2_1$
a (Å)	7.8210(10)
b (Å)	7.5940(10)
c (Å)	10.6570(10)
α (°)	90.00
β (°)	93.4520(10)
γ (°)	90.00
V (Å ³)	631.80(13)
Z	2
d (g cm ⁻³)	1.573
$F(000)$	308
μ (cm ⁻¹)	0.139
Absorption corrections	Multi-scan; 0.9727 min, 0.9754 max
Diffractometer	KappaCCD

X-ray source	Mo $K\alpha$
λ (Å)	0.71069
Monochromator	Graphite
T (K)	150.0(1)
Scan mode	Phi and omega scans
Maximum θ	27.48
HKL ranges	–10 10; –9 9; –13 13
Reflections measured	2790
Unique data	2790
Rint	0.0000
Reflections used	2680
Criterion	$I > 2\sigma I$
Refinement type	Fsqd
Hydrogen atoms	Mixed
Parameters refined	191
Reflections/parameter	14
wR2	0.0932
R1	0.0322
Flack's parameter	–0.1(5)
Weights a, b	0.0590, 0.0754
GoF	1.053
Difference peak/hole ($e \text{ \AA}^{-3}$)	0.194(0.048)/–0.179(0.048)

References

- For recent entries, see:
 - P.K. Mikhailiuk, S. Afonin, A.N. Chernega, E.B. Rusanov, M.O. Platonov, G.G. Dubinina, M. Berditsch, A.S. Ulrich, I.V. Komarov, *Angew. Chem. Int. Ed.* 45 (2006) 5659–5661;
 - B.C.J. van Esseveldt, P.W.H. Vervoort, F.L. van Delft, F.P.J.T. Rutjes, *J. Org. Chem.* 70 (2005) 1791–1795 (and references therein);
 - C.L. Jenkins, G. Lin, J. Duo, D. Rapolu, I.A. Guzei, R.T. Raines, G.R. Krow, *J. Org. Chem.* 69 (2004) 8565–8573 (and references therein);
 - P. Karoyan, S. Sagan, O. Lequin, J. Quancard, S. Lavielle, G. Chassaing, *Targets Heterocycl. Syst.* 8 (2004) 216–273;
 - J.R. Del Valle, M. Goodman, *J. Org. Chem.* 68 (2003) 3921–3933 (and references therein);
 - J. Zhang, W. Wang, C. Xiong, V.J. Hruby, *Tetrahedron Lett.* 44 (2003) 1413–1415.
- L. Moroder, C. Renner, J.J. Lopez, M. Mutter, G. Tuchscherer, in: C. Dugave (Ed.), *Cis–trans Isomerization in Biochemistry*, Wiley–VCH, Weinheim, Germany, 2006;
 - A. Flores-Ortega, A.I. Jiménez, C. Catiuela, C. Nussinov, R. Aleman, J. Casanovas, *J. Org. Chem.* 73 (2008) 3418–3427 (and references therein);
 - C. Dugave, L. Demange, *Chem. Rev.* 103 (2003) 2475–2532.
- T. Kitamoto, T. Ozawa, M. Abe, S. Marubayashi, T. Yamazaki, *J. Fluorine Chem.* 129 (2008) 286–293;
 - Y. Guo, K. Fujiwara, H. Amii, K. Uneyama, *J. Org. Chem.* 72 (2007) 8523–8526 (and references therein);
 - U.E.W. Lange, D. Baucke, W. Hornberger, H. Mack, W. Seitz, H.W. Höffken, *Bioorg. Med. Chem. Lett.* 16 (2006) 2648–2653;
 - W. Kim, R.A. McMillan, J.P. Snyder, V.P. Conticello, *J. Am. Chem. Soc.* 127 (2005) 18121–18132;
 - W. Kim, K.I. Hardcastle, V.P. Conticello, *Angew. Chem. Int. Ed.* 45 (2006) 8141–8145.
- V.P. Kukhar, V.A. Soloshonok, *Fluorine Containing Amino Acids, Synthesis and Properties*, Wiley, New York, 1995 (and references cited therein);
 - X.-L. Qiu, W.-D. Meng, F.-L. Qing, *Tetrahedron* 60 (2004) 6711–6745;
 - M. Molteni, C. Pesenti, M. Sani, A. Volonterio, M. Zanda, *J. Fluorine Chem.* 125 (2004) 1735–1743.
- G. Luppi, D. Lanci, V. Trigari, M. Garavelli, A. Garelli, C. Tomasini, *J. Org. Chem.* 68 (2003) 1982–1993 (and references therein);
 - C. Tomasini, M. Villa, *Tetrahedron Lett.* 42 (2001) 5211–5214.
- For literature concerning fluorinated pyroglutamic acids, see:
 - F.-L. Qing, X.-L. Qiu, *Synthesis of fluorinated prolines and pyroglutamic acids*, in: V.A. Soloshonok (Ed.), *Fluorine Containing Synthons*, American Chemical Society, Washington, DC, 2005, p. 562 (and references therein);
 - X.-L. Qiu, W.-D. Meng, F.-L. Qing, *Tetrahedron* 60 (2004) 5201–5206;
 - X.-L. Qiu, W.-D. Meng, F.-L. Qing, *J. Org. Chem.* 68 (2003) 3614–3617;
 - V.A. Soloshonok, C. Cai, V.J. Hruby, L. Van Meervelt, N. Mischenko, *Tetrahedron* 55 (1999) 12031–12044;
 - V.A. Soloshonok, C. Cai, V.J. Hruby, L. Van Meervelt, *Tetrahedron* 55 (1999) 12045–12058;
 - V.A. Soloshonok, D.V. Avilov, V.P. Kukhar, L. Van Meervelt, N. Mischenko, *Tetrahedron Lett.* 38 (1997) 4903–4904;
 - D. Gestmann, A.J. Laurent, E.G. Laurent, *J. Fluorine Chem.* 80 (1996) 27–30.
- α -Tfm-pyroglutamic acid has only been reported in the racemic series:
 - B. Koksche, D. Ullmann, H.-D. Jakubke, K. Burger, *J. Fluorine Chem.* 80 (1996) 53–57.
- F. Huguenot, T. Brigaud, *J. Org. Chem.* 71 (2006) 7075–7078;
 - F. Huguenot, T. Brigaud, *J. Org. Chem.* 71 (2006) 2159–2162;
 - N. Lebouvier, C. Laroche, F. Huguenot, T. Brigaud, *Tetrahedron Lett.* 43 (2002) 2827–2830;
 - T. Brigaud, G. Chaume, J. Pytkowicz, F. Huguenot, *Chim. Oggi Chem. Today* 25 (2007) 8–10.

- [9] G. Chaume, M.-C. Van severen, S. Marinkovic, T. Brigaud, *Org. Lett.* 8 (2006) 6123–6126.
- [10] The formation of similar oxazolidines has already been reported in the non-fluorinated series:
(a) C. Agami, F. Couty, B. Prince, O. Venier, *Tetrahedron Lett.* 37 (1993) 7061–7062;
(b) L.M. Harwood, K.J. Vines, M.G.B. Drew, *Synlett* (1996) 1051–1053.
- [11] In our previous report (Ref. 9) we observed that the chromatographic separation of both diastereomers of similar bicyclic CF₃-morpholinones was very easy.
- [12] The synthesis of similar bicyclic lactams has also been reported in the non fluorinated series by an alternative strategy:
L.M. Harwood, G. Hamblett, A.I. Jimenez-Dias, D.J. Watkin, *Synlett* (1997) 935–938.
- [13] R_f of (*R,S*)-7 = 0.41 (7:3 cyclohexane/ethyl acetate) and R_f of (*R,R*)-7 = 0.29 (7:3 cyclohexane/ethyl acetate).
- [14] Crystallographic data (excluding structure factors) for compound (*R,S*)-7 have been deposited with the Cambridge Crystallographic Data Centre as supplementary publication no. CCDC 695212. Copies of the data can be obtained, free of charge, on application to CCDC, 12 Union Road, Cambridge CB2 1EZ, UK (fax: + 44 1223 336033 or email: deposit@ccdc.cam.ac.uk).
- [15] G. Tang, H. Tian, D. Ma, *Tetrahedron* 60 (2004) 10547–10552.
- [16] D. Ma, G. Tang, H. Tian, G. Zou, *Tetrahedron Lett.* 40 (1999) 5753–5756.